



ISOLATION AND METABOLIC CHARACTERIZATION OF BACTERIA THAT HYDROLYZE THE UREA CONTAINED IN HUMAN URINE

María Romero*, Carmen Fajardo*, Rocío Torres**, Oscar Monroy*, Florina Ramírez*. Universidad Autónoma Metropolitana Iztapalapa, Departamento de Biotecnología*, Departamento de Hidrobiología**, México City 09340; cony549@hotmail.com

Key words: urea, urease, hydrolysis.

Introduction. Human urine containing high concentrations of urea, and other salts, urea is the main form of nitrogen in fresh urine (1). Urea, ammonium and phosphorus have value as fertilizer, may be another option compared to chemical fertilizers (2). Urease catalyzes hydrolysis of urea to yield two molecules of ammonia and carbonic acid, which establishes equilibrium between its protonated and deprotonated forms (3). The primary function of urease is to provide nitrogen as ammonia for organisms. Urease is found in seeds of plants, microorganisms and invertebrates. Regardless of the source of urease, it follows the same pattern of catalysis, primarily due to the similar amino acid sequence and the presence of two Ni²⁺ ions in the active site of the enzyme (4). The aim of this study was to isolate and characterize hydrolytic bacteria from different sources of inoculum.

Methods. Cow manure (E) and granular anaerobic sludge (L) were used as inoculum fresh. Three enrichments were performed for each inocula evaluating the urea hydrolysis. Positive cultures were isolated in Rustigian - Stuart medium (5), by mean of surface dilution technique. The pure and mixed cultures were evaluated based on the production of ammonium measured by the Nessler's method for 120 h of hydrolysis and 35°C at pH 7. Pure and mixed cultures were tested using C/N ratio of 1,3. Table 1 show the experimental design performed in this study.

Results. Three strains were isolated from granular sludge enrichment (LO₂ 10⁻¹, LO₂ 10⁻², LO₂ 10⁻³) and two from cow manure enrichment (EO₂G 10⁻²) under aerobic conditions. Table 2 show the production of ammonium as evidence of hydrolysis of urea in pure and mixed cultures. It was detected highest yields of ammonium were obtained with mixed cultures EO₂G and L which with 0,9 and 0,72 respectively, while for pure cultures low yields were obtained. The high yields of hydrolysis found in mixed cultures, may be related to the concentration of

biomass present, or with syntrophic relationships present in the inocula.

Table 1. Experimental design

Mixed cultures	Treatment	
L	Granular anaerobic sludge in anoxic conditions without additional carbon source.	
LG	Granular sludge anaerobic and anoxic conditions with glucose as carbon source.	
LO ₂	Anaerobic granular sludge in oxic conditions, without additional carbon source.	Pure culture: LO ₂ 10 ⁻¹ , LO ₂ 10 ⁻² , LO ₂ 10 ⁻³
LO ₂ G	Granular sludge anaerobic and oxic conditions with glucose as carbon source.	
EO ₂ G	Fresh cow manure, in oxic conditions with glucose as carbon source.	Pure culture: EO ₂ G 10 ⁻²

L= Granular sludge ; E= cow manure; G= glucose and O₂= Oxygen

Table 2. Production of ammonium in mixed and pure cultures as response of hydrolysis urea.

Culture	mg NH ₄ ⁺ /L			
	Initial	Final	Produced	Yield
L	568,39	4874,08	4305,68	0,72
LG	582,42	4720,43	4138,01	0,69
LO ₂	446,16	2454,94	2008,78	0,33
LO ₂ G	502,77	1706,91	1204,14	0,20
EO ₂ G	479,44	5849,70	5370,26	0,90
LO ₂ 10 ⁻¹	163,59	199,63	36,04	0,006
LO ₂ 10 ⁻²	208,86	240,53	31,67	0,005
LO ₂ 10 ⁻³	170,44	204,94	34,50	0,006
EO ₂ G 10 ⁻²	159,66	184,84	25,18	0,004

Conclusions. Mixed cultures from two sources of inoculum had a high yield in the production of ammonia in response to hydrolysis of urea.

Acknowledgements. We appreciate the support provided by CONACYT, ICyTDF and UAM (266 054).

References.

1. Larsen T., Gujer W. (1996). *Water Sci Technol.* 34(3-4):87-94.
2. Pronk W., Koné D. (2008). *Desalination.* 248(1-3): 360 – 368.
3. Mogley H., Hausinger R. (1989). *Microbiol Rev.* 53(1): 85 – 108.
4. Hanif F., Shoaib K., Saleem M., Rama N., Zaib S., Iqbal J. (2012). *SRN Pharmacol.* ID 928901.
5. Stuart C., Stratum E., Rustigian R. (1945). *J Bacteriol.* 49 (5): 437 – 444.