CULTIVATION OF MICROALGAE IN TILAPIA EFFLUENTS FOR THE PRODUCTION OF LIPIDS TO TWO LIGHTING CONDITIONS

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Introduction. Microalgae are photosynthetic unicellular organisms with higher photosynthetic efficiency higher plants for the production of biomass [1, 2]. The use of microalgae in the treatment of effluents represents one uses biotechnology environmental, being a viable alternative, due to its efficient bioconversion of solar energy, and efficiency in the use and removal of nutrients to produce biomass with high content of lipids for biodiesel production. This study evaluated lipid production from microalgae *Nannochloropsis oculata* and *Chlorella vulgaris* grown in tilapia effluent, using photobioreactors to two lighting conditions.

Methodology. Used species of microalgae N. oculata and C. vulgaris grown in Bold's Basal medium [3], an aliquot was taken of each species in its logarithmic phase as inoculum for each photobioreactor for an initial concentration of 1X10⁶ cel/ml for a total volume of 6 L. It was considered a photobioreactor with Bold's Basal medium (control) and the other with tilapia effluent (treatment) for the cultivation of each microalgae in duplicate (Table 1). Previously the physicochemical parameters of the effluent were analyzed $(N-NH_4^+, N-NO_2^- y N-NO_3^-)$. The photobioreactors were exposed to a light intensity of 4 Kluxes lamps with multi-LEDs or white light at a temperature of 25 ± 2 ° C (Fig.1). It was recorded every 24 h cell density of each species using a Neubauer Chamber, until the death of the growing phase (about 10 days). Extracted the lipid fraction by Soxhlet method [4] with chloroform/methanol (2:1), was determined the percentage and production of lipids (mg/L/d) and determined the production biomass (g/L/d).



Fig. 1. Microalgal cultures in photobioreactors: A) *N. oculata*/LEDs; B) *C.* 4. *vulgaris*/LEDs; C) *N. oculata*/white light; D) *C. vulgaris*/ white light.

Results. *N. oculata,* cultivated in tilapia effluent with lighting LEDs presented the highest cell density (1.19 x 10⁸ cel/ml) over a period of 10 days, also presented the highest production of biomass, lipid and lipid percentage (0.0410 g/L/d, 21.52 mg/L/d and 51%, respectively). *C. vulgaris,* presents a better growth in Bold's Basal medium (control) with LEDs lighting in comparison to other treatments. However, there is less production of lipids than in cultivated in tilapia effluent (Table 1).

Table 1. Average results of cell density, production of biomass, production and percentage of lipids of *N. Oculata* and *C. vulgaris* in tilapia effluent, and Bold's Basal medium (control).

Treatments			Cell density	0/ of Livide	Production of	Production of
Microalgae	Lighting conditions	Culture mediun	(Cel/ml)	% OI LIPIUS	biomass (g/L/d)	lipids (mg/L/d)
Nannochloropsis oculata	LEDs	Tilapia effluent	1.19x10 ⁸ ±2.97x10 ⁷	51.04 ± 5.83	0.0410 ± 0.0204	21.52 ± 12.75
		Bold's Basal	8.07x10 ⁷ ±2.55x10 ⁶	29.27 ± 0.33	0.0200 ± 0.0049	5.82 ± 1.42
	White Light	Tilapia effluent	6.46x10 ⁷ ±1.63x10 ⁷	39.64 ± 4.70	0.0268 ± 0.0055	9.70±0.12
		Bold's Basal	2.76x10 ⁷ ±8.13x10 ⁶	39.64 ± 4.70	0.0151 ± 0.0042	5.89 ± 0.97
Chlorella vulgaris	LEDs	Tilapia effluent	5.64x10 ⁷ ±2.76x10 ⁶	34.76 ± 2.31	0.0317 ± 0.0033	10.94 ± 0.43
		Bold's Basal	8.83x10 ⁷ ±2.83x10 ⁵	27.72 ± 5.65	0.0459 ± 0.0028	12.62 ± 1.85
	White Light	Tilapia effluent	3.44x10 ⁷ ±1.20x10 ⁶	39.38 ± 6.63	0.0446 ± 0.0079	17.29 ± 0.17
		Bold's Basal	6.54x10 ⁷ ±7.07x10 ⁵	34.94 ± 2.73	0.0384 ± 0.0016	13.40 ± 0.38

Conclusions. *N.oculata* presented higher cell density when it is cultivated with LEDs lighting and tilapia effluent. *C. vulgaris* and *N. oculata* showed increased lipid production in cultures with tilapia effluent. The use of tilapia effluent as culture medium for microalgae is more efficient in the production of lipids and biomass to the own Bold's Basal medium for both species of microalgae with LEDs lighting.

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Bibliografía.

- 1. Benemann, J.R. (1997). CO₂ mitigation with microalgae systems. Energy. Convers. Manage. 38: 475–479.
- Miao, X., Wu, Q. (2006). Biodiesel production from heterotrophic microalgal oil, Bioresour.. Technol. 97: 841–846.
- Nichols, H.W. (1973). Growth media-freshwater. In Stein, J.R. (ed.). Handbook of Phycological Methods: Culture Methods and Growth Measurements, Cambridge University Press, London. pp. 7-24.
 - Guckert, J.B, Cooksey, K.E, Jackson L.L. (1988). *Lipid solvent systems* are not equivalent for analysis of lipid classes in the microeukaryotic green alga, Chlorella. J. Microbiol. Methods. 8:139–149.