



## PARTIAL PURIFICATION OF A &R-RUTINOSIDASE FROM RHODOCOCCUS ERYTHROPOLIS DSM 44534 AND DEGRADATION EXPERIMENTS WITH DIFFERENT FLAVONOIDS AS SUBSTRATES

Schütt, D. 1,2; Martinez-Rojas, E. 2; Burkhardt, R. 1; Garbe, L.A. 1,2

1 Institute for Biotechnology, Technical University of Berlin, Seestraße 13, 13353 Berlin, Germany, 2 Researches and Teaching Institute for Brewing in Berlin (VLB), Seestraße 13, 13353 Berlin, Germany <a href="mailto:schuett@vlb-berlin.org">schuett@vlb-berlin.org</a>; <a href="mailto:denise.schuett@googlemail.com">denise.schuett@googlemail.com</a>

Key words: R. erythropolis,  $\beta$ -rutinosidase, biotransfortmation

Introduction. Glycosidases are enzymes which have various functions in nature including degradation of biomass, antibacterial defense strategies, pathogenesis mechanism and normal cellular functions. The *B*-rutinosidase is a glycosidase which hydrolyses rutin into quercetin and rutinose. It discriminate both: the sugar part and the flavonol aglycone part of its substrate. According to the literature the *B*-rutinosidase has been isolated from fungi as *Penicillium rugulosum* [1] and bacteria like *Corynebacteria sp.* [2], but for the genus *Rhodococcus* it is still unknown.

Therefore it would be interesting to elucidate the rutin catabolic pathway in *Rhodococcus erythropolis* DSM 44534.

Methods. In this study the B-rutinosidase was purified from R. erythropolis cells, which were cultivated in a modified minimal salt medium supplemented with rutin (in dimethyl sulfoxide) as carbon source at a final concentration of 100  $\mu M.$  The cultures were incubated over 10 days at 28°C and 160 rpm in a horizontal shaker. The protein was purified using 60 % ammonium sulfate precipitation at 4°C. The crude protein was dialyzed against 10 mM sodium phosphate buffer pH 6 before isoelectric focusing (IEF) with ampholyte pH-value 3-10. The activity test was performed with rutin as substrate at 37°C for 2h. The guercetin concentration was measured performance reversed phase high chromatography with diode array detector.

**Results.** 3.6 g of wet weight cells were obtained from a 3L culture. These cells were used to prepare a crude extract according to the methodology.

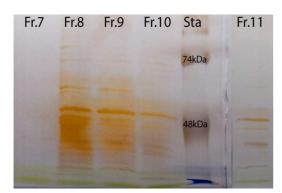


Fig. 1 10% SDS-PAGE of fraction 7-11after the IEF with prestained broad range standard

The crude protein had a concentration of 8.1 mg/mL (145 mg total protein) after the dialysis step

The biotransformation system was made with the IEF fraction 7-11 with a pH-value of 5.22, 5.52, 6.22, 6.40 and 6.72 respectively (Fig. 1). The highest level of quercentin was observed in the fraction 10 and 11 after 2 h with a concentration of 51  $\mu$ g/mL (Fig. 2). The addition of ampholyte to the sample did not transform rutin into quercentin (blank experiment).

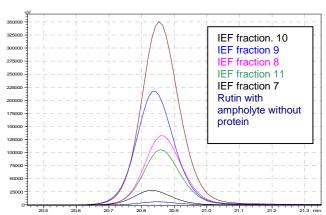


Fig. 2 HPLC-Chromatogram of rutin transformation into quercentin with IEF fraction 7-11 after 2h incubation

**Conclusion:** We have elucidated an enzyme with rutinosidase or rutinase activity from *R. erythropolis* DSM 44534. The partially pure enzyme was induced in *R. erythropolis* cells having quercentin as major product. Understanding gained from this work will be useful in studying the degradation and modification of natural products by microorganisms.

## References.

- [1] Narikawa, T; Shinoyama, H; Fujii, T., (2000). *Biosci. Biotechnol. Biochem.*, 64:1317–1319
- [2] Omori,T.; Shiozawa, K.; Sekiya, M.; Minoda, Y. (1986). *Agric. Biol. Chem. Tokyo*, 50:779–780I