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Introduction. One of the most common causes of diarrhea and even death in less than two years of developing age infants in countries is Enteropathogenic E.coli (EPEC), a facultative Gramnegative rod-shaped bacterium, member of the Enterobacteriaceae family. Its detection in foods is mainly performed by traditional culture and serological methods, which lasts more than a week. (1, 2). The aim of this work was to develop the specific detection of an EPEC strain by amplifying both bfp and eae genes coding for the bundleforming pili and the intimine proteic subunits, respectively, with multiplex-PCR (plex-PCR) (3, 4), of bacterial DNA extracted from commercial frozen burger meat in order to reduce its detection time compared with traditional culture techniques.

Methods. Primers for PCR amplification of eae and bfp genes from EPEC were designed using Genbank sequences, then were PCR amplified and sequenced. Resultant sequences were identified by comparing against the non-redundant GenBank database with the BLAST tool of the NCBI homepage. Bacterial chromosomal DNA was extracted from a pure culture of the EPEC strain inoculated in Luria Broth (37°C/24h), and from the inoculated frozen burger meat (SuKarne) enrichment culture of the strain (final concentration 1 CFU/10g), according to BAM recommended technique (5). Extracted chromosomal DNA was used as template for the 50 µL plex-PCR reaction. Reagents were according suppliers proportions to recommendation (Fermentas), the final primers concentration were 0.1 µM for eae gene and 0.2 µM for *bfp* gene, respectively. Plex-PCR conditions were: 95°C, 5 min, 1X; 95°C/1min, 56°C/1min, 72°C,1.5min, 30X; 72°C/5min, 1X; 4°C/∞. Amplicons were visualized with an agarose gel electrophoresis.

**Results.** Sequence analysis of EPEC *bfp* and *eae* correspond genes matched with some EPEC sequences in the GenBank database (data not shown). The plex-PCR products for genes *bfp* and *eae*, amplified from extracted bacterial DNA from freeze burger meat inoculated with EPEC strain and sampled at 0, 3 and 24 h resulted in two amplified bands of 2500 bp and 900 bp respectively, and were similar to those obtained with the pure strain chromosomal DNA plex-PCR (data not shown). It was observed and increase in band intensity

according with incubation time during the enrichment process (Figure 1; lines 4, 5 and 6).

M 1 2 3 4 5 6

**Figure 1.** Agarose gel electrophoresis for plex-PCR from the bacterial chromosomal DNA of frozen burger meat inoculated with EPEC 100V. Line M, 1 Kb ladder (Fermentas); Line 1, *E. coli* ATCC 11229; Line 2, EPEC *bfp* gene; Line 3, EPEC *eae* gene; Line 4, plex-PCR 0h; Line 5, plex-PCR 3 h; Line 6, plex-PCR 24 h.

**Conclusions.** A plex-PCR technique for the specific detection of an EPEC strain present in commercial frozen burger meat by amplifying the *bfp* and *eae* genes were developed. It was possible to reduce the detection time for this strain from more than a week in the traditional culture technique to less than 30 hours. EPEC strains were not detected by using this method, among the not-inoculated frozen hamburger samples analyzed,

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