



ROLE OF NCgl1221 MECHANOSENSITIVE CHANNEL IN GLUTAMATE PRODUCTION BY *Corynebacterium glutamicum*

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Key words: Corynebacterium glutamicum, mechanosensitive channel, NCgl1221

Introduction. *Corynebacterium glutamicum* was isolated in 1956 as an L-glutamate-producing bacterium by Japanese researchers. The mechanism of L-glutamate secretion by *C. glutamicum* is unique. The presence of biotin, which is required by *C. glutamicum* for growth, inhibits L-glutamate production in the culture medium, while production is induced under biotin-limited conditions. Although it has been suggested that the secretion of L-glutamate by *C. glutamicum* is mediated by a carrier system in the cytoplasmic membrane, the nature of L-glutamate exporter has long been unclear. We have recently shown that the NCgl1221 gene encoding a mechanosensitive channel homolog is involved in the mechanism of L-glutamate secretion (1). The NCgl1221 mechanosensitive channel mediates L-glutamate secretion by sensing changes in membrane tension caused by inducing treatments such as biotin limitation and penicillin addition.

The NCgl1221 protein has an N-terminal domain (1 – 286 a.a.) homologous to the *E. coli* MscS and a long C-terminal domain (287 – 533 a.a.) of unknown function. In order to investigate a role of the C-terminal domain in L-glutamate secretion, we constructed a series of C-terminally truncated mutants of NCgl1221 (2).

Methods. The PHD.htm program predicts that the C-terminal domain of NCgl1221 consists of three parts, a cytoplasmic domain (287 – 401 a.a.), a transmembrane segment (402 – 419 a.a.) and an extracytoplasmic domain (420 – 533 a.a.) (Fig. 1). Based on this prediction, we constructed following three truncated mutants; NCgl1221-419 lacking the extracytoplasmic domain, NCgl1221-401 lacking the C-terminal transmembrane segment and extracytoplasmic domain, and NCgl1221-286 lacking the whole C-terminal domain. We also constructed additional two mutants; NCgl1221-224 and NCgl1221-110, in which the N-terminal domain was truncated further (Fig. 1). Plasmids expressing these truncated mutants were introduced into the Δ NCgl1221 strain, and glutamate production was induced by biotin limitation.

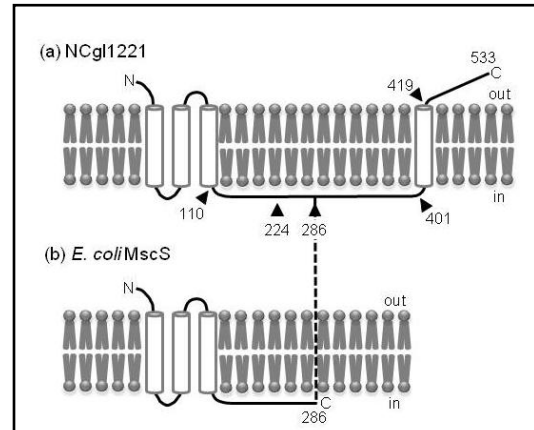


Fig.1 Membrane topology of the NCgl1221 and *E. coli* MscS. Arrow heads indicate the positions of truncation constructed in this study.

Results. The N-terminal half NCgl1221-286 homologous to the *E. coli* MscS could secrete L-glutamate (23.8 g/l) at almost comparable levels to the full length NCgl1221 (26.7 g/l). The NCgl1221-401 also secreted L-glutamate (15.8 g/l), although lower than NCgl1221-286. On the other hand, the strains expressing the shorter N-terminal domains NCgl1221-110 and NCgl1221-224 could not secrete L-glutamate (0.82 and 0.58 g/l, respectively). Interestingly, the NCgl1221-419 lacking the C-terminal extracytoplasmic domain (420 – 533 a.a.) produced L-glutamate without any inducing treatments.

Conclusions. These results clearly indicate that the N-terminal domain (1 – 286 a.a.) homologous to the *E. coli* MscS is necessary and sufficient to secrete L-glutamate in response to inducing treatments.

Acknowledgements. This work was supported in part by a grant-in-aid for Scientific Research (B) to M.W. from the Japan Society for Promotion of Science.

References.

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