



## PURIFICATION AND CHARACTERIZATION OF INVA AND INVB INVERTASES FROM ZYMOMONAS MOBILIS. EXPRESSION IN PICHIA PASTORIS BY INDUCTION WITH METHANOL

<u>Ara Itzel Pérez de los Santos-Mondragón</u>; José Alejandro Santiago-Hernández and María Eugenia Hidalgo-Lara; Departamento de Biotecnología y Bioingeniería. Centro de Investigación y de Estudios Avanzados. Mexico City 07360; araitzel@yahoo.com.mx

Key words: invA, invB genes, AOX1

**Introduction.** *Zymomonas mobilis* produce two invertases: an intracellular (INVA) and an extracellular (INVB). These enzymes have attractive features for the food industry, *e.g.*, they have less susceptibility to inhibition by sucrose than the commercial invertase from Saccharomyces cerevisie.

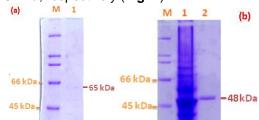
The use of modified enzymes in industry bioprocess has increased in recent years. *Pichia pastoris* has been widely used for obtaining recombinant enzyme by means of different expression systems such as the methanol-regulated using the strong promoter AOX1. *P. pastoris* has been used for the expression of proteins for both basic research and industrial processes.

The aim of this study was to express the INVA and INVB from *Z. mobilis* in *P. pastoris* under the control of the promoter AOX1 and evaluate the biochemical properties of the recombinant enzymes

**Methods.** invA and invB genes from Z. mobilis were amplified from the bacterial genomic DNA and cloned into the pPICZαB expression vector. Constructions obtained (pPICZαB-invA and pPICZαB-INVB) were transferred to P. pastoris X-33 cells by electroporation. Transformants clones were selected for Zeocin resistance, functional bioassays (1-2), PCR amplification and protein expression in P. pastoris cells by small scale fermentation and induction with 1% methanol (3). Clones with the highest invertase activity in the culture supernatant were selected for the purification of the recombinant INVA (INVA-r) and INVB (INVBinvertases. Culture supernatant was concentrated by ultrafiltration (30 kDa membranes) or precipitation  $((NH_4)_2SO_4)$ . Recombinant enzymes were purified by ion exchange chromatography, and the purified enzymes were biochemically characterized.

**Results.** The *invA* and *invB* genes from *Z. mobilis* were expressed in *P. pastoris* and the recombinant enzymes were purified from the culture supernatant of clones with the highest extracellular invertase activity.

SDS-PAGE analysis of purified INVA-r and INVB-r showed a single band of 65 kDa and 48 kDa, respectively (**Fig. 1**).



**Fig.1** SDS-PAGE analysis of purified INVA-r (a) and INVB-r (b) invertases. *P. pastoris* cells were induced with 1% methanol. (a) Line M: MW marker, Line 1: INVA-r. (b) Line M: MW marker, Line 1: Culture supernatant, Line 2: INVB-r.

Purified enzymes were biochemically characterized and a summary of results is shown in Table 1.

**Table 1.** Biochemical properties of INVA and INVB from *Z. mobilis.* pative (-n) and recombinant (-r) forms

| 2. mobilis, native ( ii) and recombinant ( ii) forms |         |               |       |
|--|---------|---------------|-------|
|  | Optimum | $V_{max}$     | $K_m$ |
|  | T (°C)  | (µmol/mg/min) | (mM)  |
| INVA-n   | 30      | n/d           | 42    |
| (Z. mobilis)   |         |               |       |
| INVA-r   | 35      | 833           | 0,042 |
| (P. pastoris)  |         |               |       |
| INVB-n   | 45      | 34            | 208   |
| (Z. mobilis)   |         |               |       |
| INVB-r   | 40      | 5000          | 0,05  |
| (P. pastoris)  |         |               |       |

**Conclusions.** The recombinant INVA and INVB invertases from *Z. mobilis*, expressed in *P. pastoris*, showed both higher catalytical activity and substrate affinity with sucrose as the substrate than those reported for native INVA and INVB invertases from *Z. mobilis*.

**Acknowledgements**. Research was funded by CINVESTAV, AIPS-M received a scholarship from CONACYT, México.

## References.

- Bochner B., Savageu M. (1977). App. Environ. Microbiol. 33 (2): pág. 434-444.
- Yanase H, Fukushi H, Ueda N, Maeda Y, Toyoda A y Tonomura K. (1998). Agric. Biol. Chem. 55 (5): pág. 1383-1390.
- Boschi S, Muniz A, Aráujo K, Marques A, Garcea R y Lee P. (2009). Arch Virol. 154: 1609- 1617: pág. 1609-1617.